# DECEMBER, 2014

a charles of the top

MASHI STA V'ATRIC No

## ANALYSIS OF BICACTIVE COMPONENTS IN ETHANOL EXTRACTS OF CLYPHAEA BREVTS LEAVES

### ANALYSIS OF BIOACTIVE COMPONENTS IN ETHANOLIC EXTRACTS OF GLYPHAEA BREVIS LEAVES

By

### DIASHI STANLEY CHIDI MATRIC NO: 12/06/2185



DEPARTMENT OF SCIENCE LABORATORY TECHNOLOGY, ABRAHAM ADESANYA POLYTECHNIC, IJEBU-IGBO, OGUN STATE, NIGERIA.

IN PARTIAL FULFILLMENT OF THE REQUIREMENT FOR THE AWARD OF NATIONAL DIPLOMA (ND) IN SCIENCE LABORATORY TECHNOLOGY.

DECEMBER, 2014

### DEDICATION

This report is dedicated to my God, the most Merciful, most Gracious, most Glorious, most Splendors and most Majesty. And to my wonderful and loving Mr. and Mrs. DIASHI

ii

. 4

. 6 .

### CERTIFICATION

I certify that this research work was carried out by DIASHI STANLEY CHIDI with Matriculation number 12/06/1285 under my supervison in the Department of Science Laboratory Technology, Abraham Adesanya Polytechnic, Ijebu Igbo, Ogun State.

111



18/55/2525

Date

Miss O.A JOKOTAGBA

Supervisor



### ACKNOWLEGDEMENT

I give thanks to almighty God for his goodness and kindness upon my life and for enabling me to complete this project. I humbly appreciate my wonderful parents who brought me into this world MR and MRS DIASHI for giving me this glorious opportunity, a solid foundation for the fulfillment of the divine success upon my life. For all their parental, financial, morally, spiritual support and for their faith in me-

supervisor Miss O.A JOKOTAGBA who inculcated in me a sense of and my prayer is that God continue bless you and perfect His work in your responsibility in the course of this project. Her tactical view, observation, suggestion and contribution were the major driving force for the successful completion of this project. I will always remember your impact in my life I pray that you will reap the fruit of your labour in good health in Jesus name and you will live to enjoy it. I give a big thanks to my wonderful, life.

because you people were part of my achievement to this level and not I also thank my family members especially my siblings Happiness, Favour, Daniel and my big sister Nelly for their trust in me and prayers and advice. My cousins the likes of Bro. Onyii Diashi, Bro. Agii Diashi, for your encouragement in my life during those I thought I have failed,

AUES ANTO

ANH. 31.1754

leaving my wonderful uncles MR M.O DIASHI, MR J.O DIASHI, MR E.O DIASHI for your financial support.

I use this medium to appreciate my Head of Department MRS OLUWABIYI for her encouragement in my life, MISS BOLANLE for her måral advice in my life and my wonderful lecturers for their wisdom "knowledge, advice they impacted in me throughout my program in this citadel of learning may God continue to bless and guide you all.

My topmost gratitude from the bottom of my heart goes to MR AKINBILE who became my parent in school made me discover who I am today, my mentor and role model for your support, advice, knowledge, understanding and teaching, May God almighty strengthen you for your support in our lives and as you continue your good work you shall reap what you have sown in Jesus name.

I also like to express deepest gratitude to this people who are like my blood brothers MR. LEKAN ADELAJA and MR. ADELEKE ADELANA you two are the best have ever met especially MR LEKAN ADELAJA you were always a part of me, and am short of words, but my prayer to God is that our friendship will not die but will continue to increase. I also thank the likes of Grace Bernard, Oluwanifemi Komolafe, Abidemi Bakare, Akintude Oluwatofurmi, Lawal Oriyomi, and my school friends

v

like Pekipuima David, Taiwo Olarenwaju, Taiwo Makinde and my entire classmate Hove you all

I finally like to appreciate my wonderful pastor in person of PASTOR OLUMIDE and PASTOR MRS OLUMIDE for their assistance in my project and their contribution in allowing me to use PLANT SCIENCES LABORATORY at Olabisi Onabanjo University may God bless you sir. I sincerely thank Sister Bunmi for her financial support, spiritual support and her advice throughout my program I will always remember you in my life forever. And my appreciation to the house of A.Y.F Pastor Taiwo, Pastor Gbolahun, Sister dupe, Sister Adeola, Pastor Daniel, and my wonderful choir mistress MRS BAKARY, Brother Damola and to all the CORAL GROUP UEBU IGBO. I love you all.

vi

-

### ABSTRACT

Plants have been an important source of medicine with qualities for thousands of years. Mainly on traditional remedies such as herbs, for their history it has been used as a nonular folk medicine. Glyphaca brevis has medicinal values: ethanolic leaf extract of this plant was analyzed using Gas Chromatography-Mass Spectrometry, Gas chromatography mass spectrometry (GC-MS) analysis revealed the presence of 16 compounds. In GC-MS analysis, some of the phytocomponents screened were Phytol, 1.2-Benzenedicarboxylic acid Oleic Acid and n-Hexadecanoic acid. The compounds were identified by comparing their retention time and peak area with that of literature and by interpretation of mass spectra. Some of the identified compounds have been reported to possess some various medicinal activities such as antioxidant, antimicrobial, anesthetic, antiseptic, antidiabetic, Hypocholesterolemic and e.t.c. The result thus was concluded that Glyphaea brevis leaves possess various bioactive compounds and it is recommended as a plant of phytopharmaceutical importances.

VIL

LYTEL HERE

. 14

### TABLE OF CONTENT

Title Page	
Pages	
Dedication	i
Certification	ii
Acknowledgment	111
Abstract	v
List of table	vi
List of Figure	vii

### CHAPTER ONE

A ....

1.0	INTRODUCTION	1
1.1	Herbalism	5
1.2	Advantage of Herbal Medicine	8
1.3	Disadvantage of herbal medicine	9
СНА	PTER TWO	
2.0	LITERATURE REVIEW	11
21	Glynhaea brevis	11

2.2 Botanical Description Habitat

2.3	Ecology and Distribution	13
2.4	Common Names	13
2.5	Taxonomy of Glyphaea brevis	13
2.6	Phytochemical Properties	14
2.7	Aims and Objectives	18
CH	APTER THREE	
3.0	MATERIALS AND METHODS	19
3.1	<sup>o</sup> Plant material	19
3.2	Methodology	19

3.2	Methodology	19
3.3	Phytochemical screening	19

### CHAPTER FOUR

4.0	RESULT AND DISCUSSION	24
4.1	RESULT	24
42	DISCUSSION	30

### CHAPTER FIVE

5.0	CONCLUSION AND RECOMMENDATION	34
5.1	Conclusion	34
5.2	Recommendation	35
REF	ERENCES	36

. ...

LIST OF TABLES.

1. Table 4.1

. . .

. . .

x

2. Table 4.2

24 26

OLYTECHNAS

### LIST OF FIGURES

Figure'2.1

Figure 4.1

. 4 .

. . .



### CHAPTER ONE

### 1.0 INTRODUCTION

Herbal, medicine has its root in prehistory making every bit as ancient tradition as farming or cooking. In the Graeco-Roman era, Hippocrates (father of medicine), Theophrastus (father of botany), Galen (originator of pharmaceutical "galenicals") and Dioseoroides were all herbalists (Moody, 2007). Also about one-quarter of the prescription drugs dispensed by community pharmacy in the United States contain at least one active ingredient derived from plants (Farnsworth and Morries, 1976).

Also in Nigeria, around 205 medicinal plant species are endemic in nature in the Northern, Western, Central and Eastern zones of the country (FEPA, 1992). Beyond the problem of trying to test herbal preparations that may contain active ingredients are the question of whether the research eventually will lead to the isolation of single active ingredient that can be packaged and sold separately. Intense debate surrounds the issue of how to conduct clinical trials of herbal medicine according to western pharmaceutical clinical standards.

Critics, say, there is an inherent problem with the single active ingredient approach preferred by pharmaceutical companies that are actively involved in herbal medicine research. It is argued that isolating a single compound may not be the most appropriate approach in situations where a plant's activity decreases on further fractionation (separation of active ingredients by using solvents) or where the plants contain two or three active ingredients, that must be taken together to produce the full effect (Chaudhury, 1992).

Beckstrom-Stember and Duke (1994) have documented several cases where synergy has been lost by using the single ingredient approach to developing drugs from plants. Other notable problems associated with herbal medicines include but not limited to how to conduct clinical trials of herbal medicine according to western pharmaceutical clinical standards, issue of dosage specifications, prominent doubts about herbal preparations such as lack of proof of their efficacy, safety, proper packaging problems, appropriateness of their degree or level of hygiene, cost of production and their level of acceptability especially among the elites in the healthcare team who continues to prescribe only orthodox medicines in hospituls and clinics (Pharmanews, 2010).

Nevertheless, the public pay high prices for orthodox medicines because the cost for experimental techniques through research and development (R and  $D_{ij}$  is, enormous. Another common perception is that orthodox medicine which is scientifically based is more reliable, safer and more

2

the man

effective. This notion may be wrong because drugs once thought to be safe are often withdrawn from the market for causing severe side effects and even intalities. The thalidomide fiasco of the 1950s and 60s was a tragic example when hundreds of women given thalidomide for early moming sickness gave birth to deformed babies.

Again, antibiotics which created false hope that modern medical science could eradicate discases caused by bacteria, ended up filling bacteria that are beneficial to human body thereby reduces the body's resistance to harmful bacteria (Bradstreet, 1998).

Recently in Migeria, the National Agency for Food Drug Administration and Control (MAFDAC) banned the use of Novalgin (a potent analgesic and ani-antipyretic agent) because of its severe side effects that led to the death of children. Although, the listory of orthodox medicine traces its root back to Hippocrates, the listory of orthodox medicine traces its medicine today is not strictly in line with the principles of the fahers of medicine (Rees and Shurer, 1996).

Orthodox medicine began over a century ago during the period of Remaissance. As at them the objective thinking of the causative theory of modern acience replaced the ecological model which had predominated for over 2000 years (Binkhe, 2004). The new paradigm is often termed the



Cartesian model being named after the French philosopher, Rene Descartes (1596-1650). This model, it was claimed, invalidated the humoral concepts of the holistic principles of Hippocrates. Galen and Ibn Sina promoted the ideology that man was separate from nature could be viewed objectively through experiment (Boussel *et al.*, 1982).

This heralded the birth of scientific or orthodox medicine. The frontiers of orthodox medicine were further broadened by Rudolph Virdow (1821-1902) who demonstrated that disease begins with changes in living cells and by Louis Pasteur (1822-1895) whose role in the development of the germ theory of infection was of key importance (Rees and Shuter, 1996; Gilbert *et al.*, 1998, Bhikha and Hao, 2000).

Under the germ theory, disease was associated with specific microorganisms. Since, then technology through research and development (R and D) had played tremendous roles in the propagation of orthodox medicine which is scientifically based and evolve along certain specifications or routes. These routes led to the manifestations of plethora of specialists in disorders of specific organs, tissue and cells such as cardiologists, dermatologists and neurologists among others.

Hence, it has been advocated that patients should be regarded as collections; of separate body parts and organ systems (Thomas, 2002).

Generally, the philosophy of orthodox medicine is exclusively based on the physical world and excludes any explanation that goes beyond this (Hammond-Tooke, 1989; Gilbert *et al.*, 1998).

For instance, health and illness are seen as a relationship between the body's components and sub-structure while the mind is considered independent of the body. The causes of disease are therefore, scientific and presented in terms of such concepts as chemical imbalance, virus replication, serum level overload and so on (Bhikha, 2004).

Technology based scientific research in herbal medicine perhaps has made some significant impact in addressing some prominent doubts about herbal preparations such as packaging problems, level of hygiene and dosage regimen. Presently, most herbal medicines sold in Nigeria.

1.1 HERBALISM

Herbalism ("herbology" or "herbal medicine"). Herbal Medicine sometimes referred to as Herbalism or Botanical Medicine, is the use of herbs for their therapeutic or medicinal value. An herb is a plant or plant part valued for its medicinal, aromatic or savory qualities. Herb plants produce and contain a variety of chemical substances that act upon the body.

Herbal medicine is the oldest form of healthcare known to mankind. Herbs had been used by all cultures throughout history. It was an integral part of the development of modern civilization. Primitive man observed and appreciated the great diversity of plants available to him. The plants provided food, clothing, shelter, and medicine. Much of the medicinal use of plants seems to have been developed through observations of wild animals, and by trial and error. As time went on, each tribe added the medicinal power of herbs in their area to its knowledgebase. They methodically collected information on herbs and developed well-defined herbal pharmaconocias.

Indeed, well into the 20th century much of the pharmacopocia of scientific medicine was derived from the herbal lore of native peoples. Many drugs commonly used today are of herbal origin. Indeed, about 25 percent of the prescription drugs dispensed in the United States contain at least one active ingredient derived from plant material. Some are made from plant extracts; others are synthesized to mimic a natural plant compound.

The World Health Organization (WHO) estimates that 4 billion people, 80 percent of the world population, presently use herbal medicine for some aspect of primary health care. Herbal medicine is a major component in all indigenous peoples' traditional medicine and a common element in

84.

Ayurvedic, homeopathic, naturopathic, traditional oriental, and Native American Indian medicine.

WHO notes that of 119 plant-derived pharmaceutical medicines, about 74 percent are used in modern medicine in ways that correlated directly with their traditional uses as plant medicines by native cultures. Major pharmaceutical companies are currently conducting extensive research on plant materials gathered from the rain forests and other places for their potential medicinal value. Substances derived from the plants remain the basis for a large proportion of the commercial medications used today for the treatment of heart disease, high blood pressure, pain, asthma, and other problems.

For example, ephedra is a herb used in Traditional Chinese Medicine for more than two thousand years to treat asthma and other respiratory problems. Ephedrine, the active ingredient in ephedra, is used in the commercial pharmaceutical preparations for the relief of asthma symptoms and other respiratory problems. It helps the patient to breather more casily.

Another example of the use of herbal preparation in modern medicine is the foxglove plant. This herb had been in use since 1775.At present, the powdered leaf of this plant is known as the cardiac stimulant digitalis to the millions of heart patients it keeps alive worldwide.

YTE H

### 1.2 ADVANTAGES OF HERBAL MEDICINE

There are a number advantages associated with using herbal medicines as opposed to pharmaceutical products. Examples include the following

1.2.1 Reduced risk of side effects: Most herbal medicines are well tolerated by the patient, with fewer unintended consequences than pharmaceutical drugs. Herbs typically have fewer side effects than traditional medicine, and may be safer to use over time.

1.2.2 Effectives with chronic conditions: Herbal medicines tend to be more effective for long-standing health complaints that don't respond well to traditional medicine. One example is the herbs and alternative remedies used to treat arthritis. Vioxx, a well-known prescription drug used to treat arthritis, was recalled due to increased risk of cardiovascular complications. Alternative treatments for arthritis, on the other hand, have few side effects. Such treatments include dietary changes like adding simple) herbs, eliminating 'vegetables from the nightshade family and reducing white sugar consumption.

1.2.3 Lower cost: Another advantage to herbal medicine is cost. Herbscost much less than prescription medications. Research, testing, and marketing add considerably to the cost of prescription medicines. Herbs tend to be inexpensive compared to drugs.

1.2.4 Widespread availability: Yet another advantage of herbal medicines are their availability. Herbs are available without a prescription. You can grow some simple herbs, such as pepermint and chamomile, at home. In some remote parts of the world, herbs may be the only treatment available to the majority of people.

### 1.3 DISADVANTAGES OF HERBAL MEDICINE

Herbs are not without disadvantages, and herbal medicine is not appropriate in all situations. These are a few of the disadvantages to consider:

1.3.1 Inappropriate for many conditions: Modern medicine treats sudden and serious illnesses and accidents much more effectively than herbal or alternative treatments. An herbalist would not be able to treat serious trauma, such as a broken leg, nor would he he able to heal an appendicitis or a heart attack as effectively as a conventional doctor using modern diagnostic tests, surgery, and drugs.

1.3.2 Lack of dosage instructions: Another disadvantage of herbal medicine is the very real risks of doing yourself harm through self-dosing with herbs. While you can argue that the same thing can happen with medications, such as accidentally overdosing on cold remedies, many herbs

9

WETT THE

do not come with instructions or package inserts. There's a very real risk of

1.3.3 Poison risk associated with wild herbs: Harvesting herbs in the wild is risky, if not foolhardy, yet some people try to identify and pick wild herbs. They run a very real risk of poisoning themselves if they don't correctly identify the herb, or if they use the wrong part of the plant.

1.3.4 Medication interactions: Herbal treatments can interact with medications. Nearly all herbs come with some warning, and many, like the herbs used for anxiety such as Valerian and SL John's Wort, can interact with prescription medication like antidepressants. It's important to discuss your medications and herbal supplements with your doctor to avoid dangerous interactions.

1.3.5 Lack of regulation: Because herbal products are not tightly regulated, consumers also run the risk of buying inferior quality herbs. The quality of herbal products may vary among batches, brands or manufacturers. This can make it much more difficult to prescribe the proper dose of an herb.

TE.

### CHAPTER TWO

### 2.0 LITERATURE REVIEW

### 2.1 GLYPHAEA BREVIS (SPRENG)

Pharmaceutical industries have come to consider traditional medicines as a source of identification of bioactive agents that can be used in the preparation of synthetic medicines. Extract from medicinal plants are sold in the partially purified or crude form for the treatment and prevention of all kinds of diseases. These herbal products lack scientific backing for the various efficacies claimed. Plants extracts are used for disease conditions such as mental disorders, diabetes, sickle cell anemia, malaria, tuberculosis and a host of other d iseases in traditional medicine throughout the world (Odugberni, 2008).

Glyphace brevis is a spreading shrub, climber or small tree up to 8m high. It is very common in undergrowth of closed forest, secondary jungle and on river-banks, lowlands to sub-mountain and wide spread in tropical. Africa (Burkill, 1985). It is valued there as vegetable (Okafor, 1990) and various therapeutic uses such as treatment of hepatitis and poisoning have been reported (Terasima and Ichikawa, 2003).

Glyphaea brevis (Spreng) which is popularly called Aloanyasi (Ibo) or Atori (Yoruba). It has been reported to have multiple physiological and

SART.

pharmacological activities. It is used in the treatment of sleeping sickness and as aphrodisiac, as an antibacterial in the treatment of eye infection and in gum cleaning. It is also reported to be effective in the treatment of impotency (Vasilea, 1969). It has carminative effects and is used as an anticonvulsant, especially in children, where it is either used singly or in combination with other herbs (Ijomah, 1996; Ogbonnia etal, 2003)

Therapeutic activities of various medicinal plants are sometimes related to their antioxidant properties (Agboret al., 2007). Therefore antioxidant activity could be accountable for the medicinal properties of *Glypheaa* brevis through a contribution to redox homeostasis.

### 2.2 BOTANNICAL DESCRPTION HABITAT

Glyphaea brevis is often in flower and fruit which makes identification easy. The stipules fall off very early but otherwise it has a classic suite of sterile characters which indicate that it is the Malvaceae. These characters are; strong stringy bark, two secondary nerves at the base of the leaf, a swollen top to the petiole and Stella hairs.

Occasionally this family can be confused with Euphorbiaceae without flowers and fruit, but the strong bark, the thinner leaves and some sliminess in the slash usually indicate Malvaceae. It is most often a small tree about 3m high, but it occasionally reaches the size of 25cm in diameter. There are usually the basal veins, on either side of the midrib.

### 2.3 \* ECOLOGY AND DISTRIBUTION

Disturbed areas on terra firma and seasonally flooded forest and riverbank.

Distributed in Guinea Bissau to Sudan, Tanzania and Angola.



Figure 2.1 Glyphaea brevis

### 2.4 COMMON NAMES

English: spreng, Yoruba: ewe Atori, Ibo: Aloanyasi

### 2.5 TAXONOMY OF Glyphaea brevis

Kingdom: <u>Plantae</u> (unranked): <u>Angiosperms</u> (unranked): <u>Eudicots</u> (unranked): <u>Rosids</u> Order: <u>Malvales</u> Family: <u>Malvaceae</u> Subfamily: <u>Grewioideae</u> Genus: *Glyphaca* 

### 2.6. PHYTOCHEMICAL PROPERTIES

2.6.1 Tannins: A tannin (also known as vegetable tannin, natural organic tannins, or sometimes tannoid, i.e. a type of biomolecule, as opposed to modern synthetic tannin) is an astringent, bitter plant polyphenolic compound that binds to and precipitates proteins and various other organic compounds including amino acids and alkaloids.

The term tannin (from *tanna*, an Old High German word for oak or fir tree, as in Tannenbaum) refers to the use of wood tannins from oak in tanning animal hides into leather; hence the words "tan\* and "tanning" for the treatment of leather. However, the term "tannin" by extension is widely applied to any large polyphenolic compound containing sufficient hydroxyls and other suitable groups (such as carboxyls) to form strong complexes with various macromolecules.

Tannins are found in leaf, bud, seed, root, and stem tissues. An example of the location of the tannins in stem tissue is that they are often found in the growth areas of trees, such as the secondary phloem and xylem and the layer between the cortex and epidermis. Tannins may help regulate the growth of these tissues.

2.6.2: Flavonoids: Flavonoids (or bioflavonoids) (from the Latin word flavus meaning yellow, their color in nature) are a class of plant secondary

metabolites. Flavonoids were referred to as Vitamin P (probably because of the effect they had on the permeability of vascular capillaries) from the mid-1930s to early 50s, but the term has since fallen out of use.

Chemically, they have the general structure of a 15-carbon skeleton, which consists of two phenyl rings (A and B) and heterocyclic ring (C). This carbon structure can be abbreviated C6-C3-C6. According to the IUPAC nomenclature.

Flavonoids are widely distributed in plants, fulfilling many functions. Flavonoids are the most important plant pigments for flower coloration, producing yellow or red/blue pigmentation in petals designed to attract pollinator animals. In higher plants, flavonoids are involved in UV filtration, symbiotic nitrogen fixation and floral pigmentation. They may also act as chemical messengers, physiological regulators, and cell cycle inhibitors.

Flavonoids secreted by the root of their host plant help *Rhizobia* in the infection stage of their symbiotic relationship with legumes like peas, beans, clover, and soy. Rhizobia living in soil are able to sense the flavonoids and this triggers the secretion of Nod factors, which in turn are recognized, by the host plant and can lead to root hair deformation and

several cellular responses such as ion fluxes and the formation of a root nodule.

In addition, some flavonoids have inhibitory activity against organisms that cause plant diseases, e.g. *Fusarium oxysporum*. Flavonoids have been shown'to have a wide rangé of biological and pharmacological activities. Examples include anti-allergic, anti-inflammatory, antioxidant, antimicrobial (antibacterial, antifungal, and antiviral, anti-cancer, and antidiarrheal activities.

2.6.3 Alkaloids: Alkaloids are a group of naturally occurring chemical compounds (natural products) that contain mostly basic nitrogen atoms. This group also includes some related compounds with neutral and even weakly acidic properties. Some synthetic compounds of similar structure are also termed alkaloids. In addition to carbon, hydrogen and nitrogen, alkaloids may also contain oxygen, sulfur and more rarely other elements such as chlorine, bromine, and phosphorus.

Alkaloids are produced by a large variety of organisms including bacteria, fungi, plants, and animals. They can be purified from crude extracts of these organisms by acid-base extraction. Alkaloids have a wide range of pharmacological activities including antimalarial (e.g. quinine), antiasthma (e.g. efficienc), anticancer (e.g. homoharringtonine), cholinomimetic (e.g. galantamine), vasodilatory (e.g. vincamine), antiarthythmic (e.g. quinidine), analgesic (e.g. morphine), antibacterial (e.g. chelerythrine), and antihyperglycemic activities (e.g. piperine).

Many have found use in traditional or modern medicine, or as starting points for drug discovery. Other alkaloids possess psychotropic (e.g. psilocin) and stimulant activities (e.g. cocaine, calfeine, nicotine), and have been used in entheogenic rituals or as recreational drugs. Alkaloids can be toxic too (e.g. atropine, tubocurarine). Although alkaloids act on a diversity of metabolic systems in humans and other animals, they almost uniformly invoke a bitter taste.

The boundary between alkaloids and other nitrogen-containing natural compounds is not clear-cut. Compounds like amino acid peptides, proteins, nucleotides, nucleic acid, amines, and antibiotics are usually not called alkaloids. Natural compounds containing nitrogen in the exocyclic position (mescaline, serotonin, dopamine, etc.) are usually attributed to amines rather than alkaloids.

2.6.4 Terpenoids: The terpenoids sometimes called isoprenoids, are a large and diverse class of naturally occurring organic chemicals similar to terpenes, derived from five-carbon isoprene units assembled and modified in thousands of ways. Most are multicyclic structures that differ from one

another not only in functional groups but also in their basic carbon skeletons. These lipids can be found in all classes of living things, and are the largest group of natural products.

Plant terpenoids are used extensively for their aromatic qualities. They play a role in traditional herbal remedies and are under investigation for antibacterial, antineoplastic, and other pharmaceutical functions. Terpenoids contribute to the scent of cucalyptus, the flavors of cinnamon, cloves, and ginger, the yellow color in sunflowers, and the red color in tomatoes. Well-known terpenoids include citral, menthol, camphor, salvinorin A in the plant *Salvia divinorum*, the cannabinoids found in cannabis, ginkgolide and bilobalide found in *Ginkgo biloba*, and the curcuminoids found in turmeric and mustard seed.

### 2.7 AIM AND OBJECTIVES

. An

The aim and objective of this project work are:

- 1) To extract active ingredients in the Glyphaca brevis leaf sample.
- 2) To carry out the phytochemical analysis of the extract.
- 3) To identify various medicinal component in the extract using Gas

Chromatography-Mass Spectrometer.

### CHAPTER THREE

### 3.0 MATERIALS AND METHODS

### 3.1 PLANT MATERIAL

The leaf of Glyphaea Brevis was obtained from owu-ikija area in ogun state, Nigeria in November 2014.

### 3.2 PREPARATION OF PLANT EXTRACTS

Fresh leaves of Glyphaea Brevis were cut and washed with water to remove all contaminants; they were air dried under room temperature and grounded to powder. The powdered leaves were extracted with ethanol using "soxhlet extractor".

### 3.3 PHYTOCHEMICAL SCREENING

Phytochemical compositions of the leaves were determined using the methods Variously described by Trease and Evans (2002).

3.3.1 Test for tannins: In the test for tannins, 0.5 g of dried powdered sample was boiled in 20 mL of water in a test tube and filtered. Few drops of 0.1% ferric chloride was added and observed for brownish green or a blue black coloration as indication of tannins.

3.3.2 Test for saponins: Approximately 2 g of powdered material was boiled in 20 mL of distilled water in a water bath and filtered. Next, 10 mL of the filtrate was mixed with 5 mL of distilled water and shaken vigorously and observed for a stable persistent froth. The frothing was mixed with 3 drops of olive oil and shaken vigorously again and then observed for the formation of emulsion as indication of saponin.

3.3.3 Test for flavonoids: A portion of the powdered material was heated with 10 mL ethyl acetate over a steam bath for 3 mins. The mixture was filtered and 4 mL of the filtrate was shaken with 1mL of dilute ammonia solution. Development of yellow coloration is an indication of the presences of flavonoids.

3.3.4 Test for steroids: In this test, 2 mL of acetic anhydride was added to 0.5 g of extract with 2mL concentrated H<sub>2</sub>SO<sub>4</sub>. The color change from violet to blue or green is indication of steroids.

3.3.5 \*Test for alkaloid: Ä measured weight of sample was dispersed in 10 % acetic acid solution in ethanol to form a ratio of 1:10 (10%). The mixture was allowed to stand for 4 hrs at 28°C. It was later filtered via what man No 42 grade of filter paper. The filtrate was concentrated to one quarter of its original volume by evaporation and treated with drop wisc



addition of concentrated aqueous NH4OH until the alkaloid was precipitated. The alkaloid precipitated was received in a weighed filter paper, washed with 1% ammonia solution dried in an oven at 80°C. Alkaloid content was Development of brownish yellow precipitate which turns intense yellow calculated and expressed as a percentage of the weight of sample analyzed. with picric acid is an indication of the presences of alkaloids.

for 1hr the mixture was filtered into 100 mL conical flask and 10 mL conical flask. 50 mL chloroform was added and shaken on a vortex mixer thoroughly for 10minutes. 3 mL of 20% NaOH was later added to develop 3.3.6 «Test for glycoside: 10 mL of extract was pipette into 250 mL pyridine, 2 mL of 2% sodium nitroprusside were added, shaken a brownish yellow color. Glycoside standard of concentrations which range from 0-5 mg/ml were prepared from 100 mg/ml stock Glycoside standard. The series of standard 0-5 mg/ml were treated similarly like sample above. The absorbance of sample as well as standard were read on a spectronic 211D Digital spectrophotometer at a wavelength of 510nm%. Trease and Evans (2002).

### 3.3.7 Cardiac glycoside

One gram of powdered sample was extracted with 10 mL of ethanol for To the filtrate 2 ml of chloroform was added and then 1ml concentrated five minutes on a steam bath and filtered. To the filtrate was added and 2-3 drops of lead acctate solution was added shaken gently and then filtered. H<sub>2</sub>SO<sub>4</sub> was carefully added to form a lower layer. A reddish-brown colour at interface was observed for cardiac glycoside.

# GAS CHROMATOGRAPHY MASS SPECTROMETER (GC-MS) ANALYSIS. 3.4

The plant powder was extracted with cthanol and analyzed using GC-MS siloxane) column (300.25mm lumdf). Helium (99.999%) was used as the carrier gas with flow rate of 1ml/min in split mode (10:1). An aliquot of 2ul of ethanol solution of sample was injected into the column with the analyzer. The data were obtained on an Elite-1 (100% Dimethyl poly injector temperature at 250° C. GC oven temperature stated at 110° C and holding for 2mins and it was raised to 200° C at the rate of 10° C/min, without holding. Holding was allowed at 280° C for 9 mins with program rate of 5° C/min. The injector and detector was tempcrature was set at  $250^{\circ}$  C and  $280^{\circ}$  C respectively. Ion source temperature was maintained at 200° C. The mass spectrum of compounds in sample was obtained by electron ionization at 70Ev and the detector was operate in scan mode from 45-459amu(atomic mass units). A scan interval of 0.5 seconds and fragment from 45 to 450 Da was maintained. The total running time was 27 minutes (sermakkani et al:2012).

. 4. 3

#### CHAPTER FOUR

## 4.0 RESULTS AND DISCUSSION

#### 4.1 RESULTS

4 8

, i .-

### Table 4.1 <u>Phytochemicals identified in the ethanolic leaf Extract of</u> *Glyphaea brevis*

Phytochemical component	Result	
Tannins	Present	
Saponins	Absent	
Flavonoid	Present	
Alkaloids	Present	
Steroids	Absent	
Glycosides	Absent	
Cardiac glycoside	Absent	
Terpenoids	Present	

1718-14

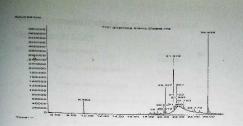


Figure 4.1: GC-MS Spectra of ethanolic extract Glyphaea brevis.

ie.



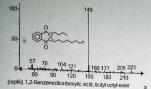
# Table 4.2: Components Identified in the ethanolic leaf extract of

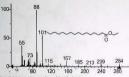
## Glyphaea brevis by GC-MS

NO	Rľ	NAMEOFCOMPOUND	MOLECULAR FORMULAR	MW
1	. 9.556	Nonane, 2, 2, 4, 4, 6; 8, 8-heptamethyl	C16H34	226.44
2	19.186	2-(Hexyloxycarbonyl)benzoic acid	C7H6O2	122.12
3	20.78	1,2-Benzenedicarboxylic acid, butyl octyl ester	C20H30Q4	334.5
4	20.216	8.8,10,10 Nonamethylcyclopentasiloxane	C <sub>10</sub> H <sub>30</sub> O <sub>5</sub> Si <sub>5</sub>	370.77
5	20.330	Hexadecanoic acid, ethyl ester	C18H26O2	284.0
6	21.377	Phytol	C29H400	296.0
7	21.503	Heptasiloxane, 1,1,3,3,5,5,7,7,9,9 ,11,11,13,13-tetradecamethyl-		
8	21.500	Trichloroacetic acid, undec-2-enyl ester	C2HCL3O2	163.39
9	21.600	Olcie Acid	$\mathrm{C}_{18}\mathrm{H}_{34}\mathrm{O}_2$	282.46
10	21.692	Pentanoic acid, 10-undecenyl ester		
11	21.755	9,12-Octadecadienoic acid, ethyl ester	$C_{20}H_{20}O_2$	280.0
12	21.806	9,12,15-Octadecatrienoic acid, methyl ester, (Z.Z.Z)-	$C_{20}\Pi_{34}O_2$	278.0
13	21.995	Octadecanoic acid, ethyl ester	C20H40O2	284.0
14	22.705	3-Isopropoxy-1,1,1,7,7,7-hexamethyl-3,5,5 tris(trimethylsiloxy) tetra siloxane	$C_{11}H_{32}\Theta_7Si_7$	577.19
15	24.170	-4 Cyclohexene-1,2dicarboxylic acid, 4- chloro-, bis(trimethylsilyl) ester		
16	25.938	Bis(2ethylhexyl) phthalate	$C_{24}H_{14}O_4$	390.56

. . .

## STRUCTURE OF IDENTIFIED COMPOUNDS





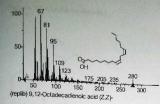
(replib) Hexadecanoic acid, ethyl ester



(mainlib) Phytol



. 4 .





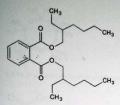
CH38,8,10,10-nonamethylcyclopentasiloxane



益



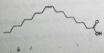
Nonane, 2,2,4,4,6,8,8-heptamethyl



Bis(2 ethylhexyl) phthalate



Heptasiloxane, 1,1,3,3,5,5,7,7,9,9 ,11,11,13,13-tetradecamethyl



Oleic Acid



8,8,10,10-nonamethylcyclopentasiloxane



#### 2-(Hexyloxycarbonyl)benzoic acid

#### 4.2 DISCUSSION

Medicinal plants are used in traditional treatment to cure variety of diseases. In the last few decades there has been an exponential growth in the field of herbal medicine. Thus, for the above mentioned reason and bearing in mind its medicinal importance, the plant species *Glyphaea brevis* were selected to analyze by GC-MS technique and explore the major and minor phyto-constituent present in the respective

plant species. The preliminary phytochemical screening shows the presence of tannins, flavonoid, alkaloids and tepemoids.

20

The GC-MS analysis of Gyphaea brevis revealed the presences of some compounds. The identified compounds possess so many biological properties. Phytol is one among the seventeen compounds of the present study. Similarly Maria Jancy Rani et al. (2011) observed the presence of phytol in the leaves of Lantana camara and Sridharan et al. (2011) in Mimosa pudicu leaves. Similar result was also observed in the leaves of Lantana camara (Sathish kumar and Manimegalai, 2008). Phytol was observed to have antibacterial activities against Staphylococcous aureus by causing damage to cell membranes as a result there is a leakage of potassium ions from bacterial cells (Inoue et al., 2005).4 Moreover, Hexadecanoic acid, ethyl ester determined at (R/T 20.330), also serve as antioxidant, Flavour, ['Hypocholesterolemic, Nematicide, Pesticid e, Lubricant, Antiandrogenic, Haemolytic, 5-Alpha reductase inhibitor as discovered by sermakanni M. et al; (2012). Furthermore, 9,12 Octadecadienoie acid determined at (R/T 21.600) serves as anti-inflammatory, Nematicide, Insectifuge, Hypocholesterolemie, Cancer preventive, Hepatoprotective, Antihistaminic, Antiacne, Antiarthritic, Antieczemie, as determined by sermakanni M. et al; (2012).

Nevertheless, 9,12,15-Octadecatrienoic acids methyl ester determined at ( R/T 21.806) serves as Anti-inflammatory, Hypocholesterolemic, Cancer preventive, Hepatoprotective, Nematicide, Insectifuge Antihistaminic, Antiarthritic, Anticeronary, Anticezemic Antiacne, 5-Alpha reductase inhibitor Antiandrogenic, as described by senthamarai Selvi. V et al; (2012). While pentadecanoic acid is used as an Antioxidant. And 1, 2-Benzenedicarboxylic acid, butyl octyl ester is an plasticizer compound which act as a Antimicrobial, antifouling Substances.

However Trichloroacetie acid, It is used in biochemistry for the precipitation of macromolecules, such as proteins, DNA, and RNA. Solutions containing trichloroacetic acid as an ingredient are used for cosmetic treatments, such as chemical peels, tattoo removal, and the treatment of warts, including genital warts. It can kill normal cells as well. It is considered safe for use for this purpose during pregnancy.

Bis(2 ethylhexyl) Phthalate is widely used as a plasticizer in manufacturing of articles made of PVC. It is also used as a hydraulic fluid

and as a dielectric fluid in capacitors. Bis (2 ethylhexyl) Phthalate also finds use as a solvent in glow sticks.

Finally, Benzoic acid which is an aromatic compound serves as a biological uses for Allergenic, anesthetic, antibacterial, anticancer, antimutagenic, antipeptic, antiseptic, antispasmodic, antitumor, candidicide, flavour, insecticide, nematicide, pesticide, sedative, termiticide, tyrosinase inhibitor.

. . .

**a** 2 , 12

-

### CHAPTER FIVE

## 5.0 CONCLUSION AND RECOMMENDATION

#### 5.1 CONCLUSION

The source of many plants (herbs and spices) can often be identified from the peak pattern of the chromatograms obtained directly from headspace analysis. GC-MS method is a direct and fast analytical approach for identification of terpenoids and steroids and only few grams of plant material is required.

The importance of the study is due to the biological activity of some of these compounds. The present study, which reveals the presence of components in *Glyphaea brevis* suggests that the contribution of these compounds from the extracts will be valuable for treatment of ailments by traditional practitioners.

Therefore, *Glyphea brevis* leaves can be used as antimicrobial, antiparasitic insecticide, nematicide, pesticide, antiandrogenic, hypocholesterolemic, antioxidant, cancer preventive, anticoronary, antiarthritic, hepatoprotective, neuroactive, analgestic and anesthetic, flavour and allergenic.

## 5.2 RECOMMENDATION

À

The phytoconstituents of *Glyphaea brevis* possess various potent bioactive compounds and is recommended as a plant of phytopharmaceutical importance. Therefore, further studies on isolation and identification of individual constituent are very much needed. It is also timely to explore its pharmacological values at the molecular level with the help of various biotechnological techniques in future.

#### REFERENCES

34-7.

Agbor G, Kuate D, Oben J (2007). Medicinal plants can be good sources of antioxidants: Case study in Cameroon. Pakistan J. Biol. Sci., 10(4): 537-544

Amusa, N.A, Adegbite, A.A., Muhammed, S. andBaiyewu, R. A., Yam diseases and its management inNigeria. Afric. J. Biotech., 2(12), 2003, 497-502.

Amusa, N.A., and Baiyewu, R.A. Storage andmarket disease of yam tubers in southwestern Nigeria.Ogun. J. Agric. Res. 2, 1999, 35-39.

Amadioha, A.C., and Obi, V.J. Control of anthracnose disease of Cowpea by Cymbopogon citrates and Ocimum gratissimum. Acta phytopathol. Entomol. Hungerica., 34(1-2), 1999, 83-89.

Aboagye-Nuamah, F., Offei, S.K., Cornelius, E.W. and Bancroft, R.D. Severity of spoilage storage rots of white yam (*Dioscorea rotundata* Poir.). Annals Appl. Biol., 147 (2), 2005, 183-190.

Alekhyå V, Deepan T, Shaktiprasanna S, Dhanaraju MD. Preliminary phytochemical screening and evaluation of *in vitro* antioxidant activity of *Anthocephalous codumba* by using solvent extracts. Eur J Biol Sci 2013;5(1):

Acharyya S, Dash DK, Mondal S, Dash SK. Studies on glucose lowering efficacy the *Anthocephalus cadamba* (Roxb.) Miq roots. Int J Pharm Bio Sci 2010;2(1):1-9.

Babalola O.O, Areola J. O (2010). Interactive roles of terpenoid extract from the leaves of *Azadirachta indica* (A. Juss) on lead induced toxicity in pregnant rabbits. J. Med. Plant Res.,4(12): 1102-1107

Coyne, D.L., Tchabi, A., Baimey, H.,Labuschagne, N., and Rotifa, I. Distribution and prevalence of nematodes (*Scutellonema bradys* and *Meloidogyne spp.*) on marketed yam (*Dioscorea spp.*) in West Africa. Field Crop Res., 96(1), 2006, 142-150

Chaman L, Verma LR. Use of certain bioproducts for insect pest control. Indian J Traditional Knowledge 2006;5(1):79-82.

Dubey A, Nayak S, Goupale DC. A review on phytochemical, pharmacological and toxicological studies on *Neolamarckia cadamba*. Der Pharmacia Letter 2011;3(1):45-54.

Domsch, K.H., Gams, W., and Anderson, V. Compendium of soil fungi, 1980, Academic Press, London

Dake JA. Phytochemical and Ethnobotanical Databases. (http://www.ars-grin. gov/duke/chem-activities.html); 2007.

Dakam W, Oben JE, Ngogang JY (2008). Antioxidant activity and extractability of phenolic compounds from medicinal plants: A study of *Glyphaea brevis*. Pharmacologyonline 3:

708-718.

Esimone, C.O., Adikwu, M.U. and Okonta, J.M.Preliminary antimicrobial screening of the ethanol extract from the lichen *Usnea subfloridans* (L.). J. Pharm. Res. Dev., 3(2), 1998, 99-102.

Edeoga HO, Okwu DE, Mbaebie BO (2005). Phytochemical constituents of some Nigerian medicinal plants. Afr. J.Biotechnol., 4(7): 685-688. FAO. Food and Agriculture Organizationproduction year book, 2004, Rome.

Fiagan Y.S. Amélioration du stockage et de la conservation. Des ignames. evaluation technique et économique: Expérience du Bénin. Proceedings of the Workshop on the African experience on post-harvestt technology development, Accra (Ghana), 4-8 July 1994.Accra, pp. 23-25.

THEMININIZ

Hegde K, Thakker SP, Joshi AP, Shastry CS, Chandrashekhar KS. Anticonvulsant activity of Carissa carandas Linn. root extract in experimental mice. Tropical J Pharm Res 2009;8:117-25

Ho WS, Pang SL, Julaihi A. Identification and analysis of expressed sequence tags present in xylem tissues of kelampayan (*Neolamarckia cadamba* (Roxb.) Bosser)<sup>10</sup>. Physiol Mol Biol Plants 2014;20(3):393-7.

Himanshu GSK, Nandanwar JR, Vinod KS. Phytochemical screening on the stem bark of *Anthocephalus cadamba* (Roxb.) Miq. Int J Pharm Sci Res 2010;1(7):108-15.

IITA. Annual Report, United Kingdom, 2009, pp.2-37.

Kumar V, Mahdi F, Chander R, Singh R, Mahdi AA, Khanna AA, et al. Hypolipidemic and antioxidant activity of *Anthocephalus indicus* (Kadam) root extract. Indian J Biochemistry Biophysics 2010;47:104-9.

Lai PS, Ho WS, Pang SL. Development, characterization and cross-species transferability of expressed sequence tag-simple sequence repeat (EST-SSR) markers derived from kelampayan tree transcriptome. Biotechnol 2013;12(6):225-35. Madhu C, Sharma U, Kumar N, Singh B, Satwinderjeet K. Antioxidant activity and identification of bioactive compounds from leaves of Anthocephalus cadambaby ultra-performance liquid chromatography/electrospray ionization quadrupole time of flight mass spectrometry. Asian Pacific J Tropical Medicine 2012;5(12):977-85.

Nweze, E.I., Okafor, J.I. and Njoku, O. Antimicrobial activities of methanol extracts of *Trema guineensis* (Schumm and Thorn) and *Morinda lucida* Benth used in Nigerian herbal medicinal practices. J. Biol. Res. Biotechnol., 2 (1), 2004, 39-46.

Neuwinger, H.D., (1996). African ethnobotany: poisons and drugs. Chapman & Hall, London, United Kingdom, 941 pp.

1

. 3

Neuwinger, H.D., (2000). African traditional medicine: a dictionary of plant use and applications. Medpharm Scientific, Stutgart, Germany. 589 pp.

Njokwu P.C. and M.I.Akumefula(2007); phytochemical and Nutrients Evaluation of spondias mombin leaves. Pakistan Journal of Nutrition, 6(6);613-615.

40

SOLVTE HNS

Ide AN I WI

Okigbo, R. N. Fungi associated with peels ofpostharvest yams in storage. Glob. J. Pure. Appl. Sci., 9,2003, 19-23

Okigbo, R.N., and Nmeka, I.A. Control of yam tuber rot with leaf extracts of *Xylopia aethiopica* and *Zingiber officinale*. Afr. J. Biotech., 4 (8), 2005, 804-807.

Ogbulie, J.N., Uwaezuoke, J.C. and Ogiebor, S.I. Introductory Microbiology Practicals, 2nd Ed. Concave Publishers. Nigeria, 2001, pp. 95 - 113.

Osadebe, P.O. and Ukwue, S.E. A comparative study of the phytochemical and antimicrobial properties of the eastern Nigerian specie of African mistletoe (*Loranthus micranthus*) sourced from different host trees. J. Biol. Res. And Biotech., 2(1), 2004, 18 – 23.

Oduro, A.C. (2000). Survey of tree species associated with cocoa cultivation in Osino District of Ghana and vegetative propagation of three of them. M.Phil. Crop Science degree

Ohuabunwa,S.I.(1998). Modern Herbal Medicinal Products. Proceedings of 1<sup>st</sup> international workshop on Herbal medicinal products, Nov.22-24, University of Ibadan, standardization and utilization of Herbal Medicines,

pp:1-24

Osemene, K.P; M.O. Illori and A.A.Elujoba.(2011). Generation and acceptability of herbal medicines research and development output in Nigeria. Res.J.pharm.Technol;4:121-130

Patel HJ, Sarra J, Caruso F, Rossi M, Doshi U and Stephani RA.(2006). Synthesis and anticonvulsant activity of new N-1',N-3'-disubstituted-2'H,3H,5'H-spiro-(2-benzofuran-1,4'-imidazolidine)-2',3,5'-triones. Bioorg. Med. Chem. Lett. 2006; 16: 4644-4647

Pharmanews ,( 2006). Innovations and R and D activities in the Nigerian manufacturing pharmaccutical industries. Editorial comment. Nig. Foremost Health J:14:34-41.

Pharmanews ,( 2010), The use of herbal medicines in hospitals in Nigeria. Foremost Health J;10:2-2

Prajapati N, Purohit D, Sharma SS, Kumar AK. A handbook of medicinal plants: a complete source book. Agrobios (india) publisher, 2007.

Patel DA, Patel YK, Shah PB. Pharmacognostical study of Neolamarckia cadamba (Roxb.) Bosser bark. Int Res J Pharm 2012;3(6):120-21.

42

MANUNIAUI

the.

Radl S, Hezky P, Urbankova J, Vachal P,(2000) Krejef I. Synthesis and Analgesic activity of some 1-Benzofurans, 1-Benzothiophenes and Indoles. Collect. Czech. Chem. Commun. 65: 280-296.

a.

Reddy KA, Lohray BB, Bhushan V, Bajji AC, Reddy KV, Reddy PR et al.(1999) Novel Antidiabetic and Hypolipidemic Agents. 3. Benzofuran-Containing Thiazolidinediones. J. Med. Chem. 42: 1927-1940. Rees, B. and P. Shuter,(1996).Medicine Through Time. Heinemann, London. ISBN-13:97804353309220

Reed J.D;Horvath P.J; Allen M.S; Van Soest P.J.(1985). Gravimetric determination of soluble phenolics including tannins from leaves by precipitation with trivalent ylterblum.J.Sci.Food Agrie;36:255-261 Gravimetric method with pvp (makkar et al;1998). http://www.ansci.cornell.edu/plants/toxic agents/tannin.html.

Ross Jr, RR; Conway,GF (1970),Hemorrhagic cystitis following accidental overdose of methenamine mandelate. American Journal of disease of children. 119(1):86-87.PMID5410299

Sermakkani and Thangapandien (2012) GC-MS analysis of cassia italicia leaf methanol extract , Asian Journal of pharmaceutical and clinical

Research. Vol 5(2),pg 641.

4 4

43

TIMPIC STREET

S. Gopalakrishnan(2011) GC-MS analysis of some bioactive constituents of MUSSAEND FRONDOSA LINN, International Journal of pharma & Bio sciences. Vol 2(1) pg 313-320

\* \*

Sobota, A.E(1984), Inhibition of Bacterial Adherence by cranberg Juice: Potential use for the Treatment of Urinary Tract Infections. Journal of Urology:1013-1016.

Sofowora A (2008). Medicinal plants and traditional medicine in Africa, 3rd Edition John Wiley and sons Ltd. New York, pp.199-202. Thomas , p.( 2002). What works, What Doesn't : The Guide to Alternate Health care. Gill and Macmillan. Ltd; Dublin.

Trease G.E, W.C. Evans, (2002), Pharmacological Activities. Pharmacognosy 15<sup>th</sup> edn, saunders publishers, London, pp 42-44.

Tchín BL, Ho WS, Pang SL, Ismail J. Association genetics of the cinnamyl alcohol dehydrogenase (CAD) and cinnamate 4-hydroxylase (C4H) genes with basic wood density in *Neolamarckia cadamba*. Biotechnology 2012;11(6):307-17. Tiong SY, Ho WS, Pang SL, Ismail J. Nucleotide diversity and association genetics of xyloglucan endotransglycosylase/ hydrolase (XTH) and cellulose synthase (CesA) genes in *Neolamarckia cadamba*. J Biol Sci 2014;14(4):267-375

WHO<sub>3</sub>(1996). Final report of the seminar on the use of medicinal plants in health care .WHO (WPRO publication), TokyodjggggjS

45

STREATION/2019-22

4 2

. 4. 4